TECHNICAL NOTE

Quenching of Fluorescence with Temperature

TN_P27; 15 Feb 2016, Georgios Arnaoutakis, Dirk Nather

Introduction

Reliable fluorescence standards and stable fluorescent probes for bio-analytics and chemistry are as important as sensitive indicator fluorophores that utilise the outstanding property of fluorescence of being highly sensitive to the fluorophore's micro-environment.

One of the observable fluorescence parameters, the fluorescence intensity, can be affected by two distinctively different quenching processes: dynamic or static quenching.¹ The fact that a fluorescent molecule can show either dynamic, or static quenching, or a combination of both, limits the chances for this molecule to be used as a standard, where often insensitivity is required over a large range of parameters such like temperature, pH, polarity, concentration, etc. On the other hand, the property of quenching, in particular that of dynamic quenching, can be used as a sensitive indicator in biological or chemical experiments and assays.

The xanthene dye Rhodamine, has been extensively used for both fluorescence standard^{2,3} and probe purposes.^{4,5} However, there is a variety of different forms of Rhodamines, and these variants can be distinctively different in their response to temperature, polarity and pH.

In this technical note we use the Spectrofluorometer FS5 and the temperature controlled sample module SC-25 to show the strong temperature dependence of the fluorescence emission spectra of Rhodamine-B. Using the extended capabilities of FS5-TCSPC we also record, for the same set of temperatures, the fluorescence lifetimes. We can show that temperature dependence of Rhodamine B is exclusively caused by dynamic quenching.



Figure 1: FS5 Spectrofluorometer

Methods and Materials

Spectral temperature quenching experiments were ran with the FS5 Spectrofluorometer in standard configuration with a 150 W xenon lamp and a single photon counting photomultiplier (PMT) detector (Hamamatsu, R928P).

Time resolved fluorescence measurements were made by Time-Correlated Single Photon Counting (TCSPC) with 478 nm excitation provided by a pulsed picosecond diode laser (EPL-485). For these fluorescence decay measurements an additional 515 nm long-pass filter was used to block residual





excitation from being detected.

A thermoelectrically cooled/heated cuvette holder cassette (SC-25) with integrated controller (TC-125) was used to set and monitor the temperature of the sample. The temperature was monitored for 5 min before each spectral and time-resolved measurement, to allow the sample to thermally stabilise. The entire measurement sequence from 5oC to 70oC in intervals of 5oC (with stabilisation times and automated measurement stop conditions) is fully computer controlled by means of the spectrometer's operating software, Fluoracle®. This applies for both spectral and lifetime measurements.

Fluoracle® also contains standard reconvolution fit analysis software. Fluorescence lifetimes *T* were obtained by fitting to a single exponential model equation $I(t)=A+Be^{(-t/T)}$. Solutions of Rhodamine-B (Sigma-Aldrich, 234141) were prepared in water (Sigma-Aldrich, 95283) in 1 cm quartz cuvettes to an OD_{525nm}=0.2 to keep aggregation effects to a minimum.⁶

Results and Discussion

The temperature dependent emission spectra of Rhodamine-B in water are displayed in Figure 1 from 16 different temperatures between 5°C to 80°C. The emission intensity increases as temperature decreases. As the absorbance stays constant over this temperature range (results not shown) we conclude that this is consistent with a change in the fluorescence quantum yield. Temperature dependent changes of Rhodamine-B in a different solvent have also been shown in literature² and explained with mobility changes of the molecule's diethylamino groups.⁷ The temperature also affects the radiative lifetime as shown in Figure 2 for the same temperature range.



Figure 1: Emission spectra of Rhodamine-B in H₂O for temperatures 5°C - 80°C. The parameters of the measurements were λ_{exc} =525 nm, $\Delta\lambda_{exc}$ = $\Delta\lambda_{em}$ =1 nm, step=1 nm, dwell=1 s, t_{stab}=5 min.

TECHNICAL NOTE

Quenching of Fluorescence with Temperature





Figure 2: Radiative decays of Rhodamine-B in H2O from 5°C - 80°C. Measurement parameters were λ_{exc} =478 nm, λ_{exc} =575 nm, $\Delta\lambda_{em}$ =5 nm, t_{cal} =0.024 ns, t_{stab} =5 min.

The integrated fluorescence intensities of the measurements shown in Figure 1 and the fluorescence lifetimes, T, obtained by numerical reconvolution from the measurements shown in Figure 2 are summarised in Figure 3, below.



Figure 3: Integrated intensity and lifetime of Rhodamine-B as a function of temperature.

The perfect overlay demonstrates that the change in the fluorescence lifetime is the only cause for the change of the fluorescence intensity. This clearly proves the pure dynamic quenching of the Rhodamine-B fluorescence by temperature increase. At high temperatures, diffusion occurs faster, which leads to greater collisional quenching.¹ The results can be further used to perform additional calculations, such as monomer-dimer binding constants.⁸

Conclusion

Quenching and temperature dependence of fluorescence in Rhodamine-B has been presented in this technical note. The effect of temperature in the fluorescence and lifetime was readily monitored with the aid of a temperature controlled SC-25 cassette via steady-state and timeresolved techniques. The strong effect of temperature was demonstrated and is associated with dynamic quenching of the fluorescence by temperature related mobility changes of groups within the Rhodamine-B molecule structure. The presented measurements can be further used for estimating the binding constants of new fluorophores.

References

- Albrecht, C. Joseph R. Lakowicz: Principles of fluorescence spectroscopy, 3rd Edition. 390, (2008). pp. 278-280
- [2] Karstens, T. & Kobs, K. Rhodamine B and rhodamine 101 as reference substances for fluorescence quantum yield measurements. J. Phys. Chem. 84, 1871–1872 (1980).
- [3] Kubin, R. F. & Fletcher, A. N. Fluorescence quantum yields of some rhodamine dyes. *Journal of Luminescence* 27, 455–462 (1982).
- [4] Miao, X., Wilson, B. K., Pun, S. H. & Lin, L. Y. Optical manipulation of micron/submicron sized particles and biomolecules through plasmon ics. Opt. Express, OE 16, 13517–13525 (2008).
- [5] Zhang, X.-F., Zhang, Y. & Liu, L. Fluorescence lifetimes and quantum yields of ten rhodamine derivatives: Structural effect on emission mechanism in different solvents. *Journal of Luminescence* **145**, 448–453 (2014).
- [6] Selwyn, J. E. & Steinfeld, J. I. Aggregation of equilibriums of xanthene dyes. J. Phys. Chem. **76**, 762–774 (1972).
- [7] Lopez Arbeloa, F., Lopez Arbeloa, T., Tapia Estevez, M. J. & Lopez Arbeloa, I. Photophysics of rhodamines: molecular structure and solvent effects. J. Phys. Chem. 95, 2203–2208 (1991).
- [8] Kemnitz, K. & Yoshihara, K. Entropy-driven dimerization of xanthene dyes in nonpolar solution and temperature-dependent fluorescence decay of dimers. J. Phys. Chem. 95, 6095–6104 (1991).

For more information, please contact:

T: +44 (0) 1506 425 300 E: sales@edinst.com F: +44 (0) 1506 425 320 W: www.edinst.com



Words and logos marked with © or TM are registered trademarks or trademarks owned by EDINBURGH INSTRUMENTS Limited. Other brands and names mentioned herein may be the trademarks of their respective owners. Neither the whole nor any part of the information contained in, or the product described in, this document may be adapted or reproduced in any material form except with the prior written permission of the copyright holder.